

Eureka Herba Indonesia

Journal Homepage: https://eurekabiomedical.com/index.php/EHI

Potential Combination of Tinospora crispa, Andrographis paniculata, Cinnamomum burmanii, Syzygium polyanthum, and Momordica charantia Extracts Against Intake of Glucose in Muscle Rats-Induced Diabetes Mellitus Rachmat Hidayat^{1*}, Patricia Wulandari²

- ¹ Department of Biology, Faculty of Medicine, Universitas Sriwijaya, Palembang, Indonesia
- ² Cattleya Mental Health Center, Palembang, Indonesia

ARTICLE INFO

Keywords:

Tinospora crispa Andrographis paniculata Cinnamomum burmanii Syzygium polyanthum Momordica charantia

*Corresponding author:

Rachmat Hidavat

E-mail address:

dr.rachmat.hidayat@gmail.com

All authors have reviewed and approved the final version of the manuscript.

https://doi.org/10.37275/ehi.v2i1.14

1. Introduction

Diabetes mellitus is a chronic condition that disturbs the body's blood sugar regulation. This disorder is characterized by a decrease in the ability of body cells to intake glucose into cells. Due to the failure of cells in glucose intake, glucose buildup occurs in the extracellular, namely in the plasma. The buildup of glucose in the plasma causes an increase in blood glucose levels in the plasma so that it will interfere with blood flow, which results in blood viscosity and decreased blood flow to various cells and tissues. Impaired blood flow to various cells and tissues leads to a decrease in the supply of oxygen and nutrients to cells and tissues resulting in damage and cell death which leads to decreased tissue

ABSTRACT

Diabetes mellitus is a chronic condition that disturbs the body's blood sugar regulation. This disorder is characterized by a decrease in the ability of body cells to intake glucose into cells. Due to the failure of cells in glucose intake, glucose buildup occurs in the extracellular, namely in the plasma. These medicinal plants contain quite varied secondary metabolites, and these plants are rich in flavonoids, alkaloids, terpenes, and glycosides. The content of these secondary metabolite compounds is believed to be rich in antioxidant effects so that it has the effect of being able to suppress various oxidative stress conditions that cause damage to various organs due to blood sugar dysregulation. The effect of *Tinospora crispa*, *Andrographis paniculata*, *Cinnamomum burmanii*, *Syzygium polyanthum* and *Momordica charantia* extract on blood sugar levels and GLUT4 protein expression in muscle tissue shows the potential of the test extract's ability to improve blood glucose intake to cells so as to maintain blood sugar regulation.

performance.1-3

Tinospora crispa, Andrographis paniculata, Cinnamomum burmanii, Syzygium polyanthum and Momordica charantia are one of the most common plants in Indonesia. This plant is often found in various regions in Indonesia, where these plants are often in the form of shrubs or wild plants that grow in yards or plantations. These medicinal plants contain quite varied secondary metabolites, and these plants are rich in flavonoids, alkaloids, terpenes, and glycosides. The content of these secondary metabolite compounds is believed to be rich in antioxidant effects so that it has the effect of being able to suppress various oxidative stress conditions that cause damage to various organs due to blood sugar dysregulation.4-6

This study aims to assess the effects of the extracts of *Tinospora crispa*, *Andrographis paniculata*, *Cinamomum burmanii*, *Syzygium polyanthum*, and *Momordica charantia* on blood sugar levels and GLUT4 protein expression in muscle tissue which shows the potential of the test extract's ability to improve blood glucose intake to cells so as to maintain sugar regulation blood.

2. Methods

Animal model

A total of 30 white rats (Rattus norvegicus) Wistar strain was obtained from the Eureka Research Laboratory (Palembang, Indonesia) weighing between 200 - 250 grams. All experimental animals were kept in cages under controlled conditions of 12 hours of the light-dark cycle, temperature 22 ± 1°C and humidity 40-60%, and given ad libitum food. The research treatments and procedures have received approval from the medical research ethics committee of the Faculty of Medicine, Sriwijaya University (No. 243 / kptfkunsri-rsmh / 2020).

Herbal combination extraction preparation

Herbal Simplicia was obtained from the Tawangmangu Herbal Research Center, Karanganyar, Indonesia. The process of extracting sambiloto is carried out by maceration in which 500 grams of Simplicia are macerated with 96% ethanol for 72 hours. Next, do the separation between the pulp and the macerate. The macerate was then evaporated with a rotary evaporator (Shimadzu) in order to obtain a thick extract, herbal combination extract (EKH).

Animal model of diabetes mellitus

After 1 week of adaptation, the mice were randomly divided into the following six groups, each containing 5 animals: Normal control group, diabetes group (negative control), diabetes + metformin group (Met; 45 mg / kg), Diabetes + ES group (50 mg / kg), diabetes + ES group (100 mg / kg) and diabetes + ES group

(200 mg / kg). Induction of diabetes was done by injecting alloxan at a dose of 110 mg / kgBW intraperitoneally, then white mice were given 10% glucose to drink. The positive control group was treated with metformin (Dexa Medica, Indonesia) for 14 days. In the treatment group, the extract of sambiloto was carried out for 14 days. The mice were anesthetized by injecting 10% Chloral Hydrate (3.5 ml/kg) intraperitoneally. Rats were sacrificed by intraperitoneal injection of 10% chlorine hydrate, then blood serum was taken through the orbital vein and the femral muscle was taken from the thigh of the white rat. The serum was then centrifuged at 10.000 rpm for 10 minutes, at 25°C and the supernatant was stored at -20°C for analysis of blood sugar levels using the spectrophotometer method (Biorad). Meanwhile, the muscle tissue was evacuated, some of which were homogenized and centrifuged to obtain a supernatant and put it in a later RNA solution (Sigma Aldrich) and stored at -20°C, for ELISA examination of GLUT4 protein.

Enzyme-linked immunosorbent assays (ELISA) GLUT 4

GLUT4 levels in joint synovial fluid were examined with Rat ELISA GLUT4 (Cloud Clone), based on the protocol contained in the manufacturer's protocols. Briefly, 50 µl of standard diluent or serum samples were added to the well coated with anti-GLUT4 and incubated at 37°C for 30 minutes. After the plates were washed, 100 µl of the biotinylated antibody solution was added and incubated for 30 minutes at 37°C. After washing three times, 50 ul of the avidin-peroxidase complex solution was added and incubated for 15 minutes at 37°C. After washing, 50 ul of tetramethylbenzidine color solution was added and incubated in the dark for 15 minutes at 37°C. Finally, 50 ul stop solution was added to stop the reaction, and the optical density (OD) was measured using an ELISA reader (Biorad), a wavelength of 450 nm.

Phytochemical test

The extract was analyzed for phytochemical screening which included tannins, alkaloids, flavonoids, quinones, saponins, and steroids/triterpenoids. The extract was separated using TLC as a stationary phase in the form of silica gel GF254 and the mobile phase in the form of n-hexane: chloroform: ethyl acetate (2: 5: 5).

Statistical analysis

All data were presented as mean ± standard deviation and all statistical analyzes were performed with the SPSS 25 (IBM) program. One-way ANOVA followed by post hc analysis was carried out to assess the difference in mean expression levels of each protein. P <0.05 was determined as an indication that there was a significant difference in mean levels.

3. Results and Discussion

Table 1 shows the potential of the extract on blood sugar levels of White Rats. Alloxan-induced white rats showed a very significant increase in blood sugar levels, where the use of the drug metformin was able to reduce blood sugar levels significantly even though they had not reached the target blood glucose target, which was less than 200 mg / dL. The treatment with the combined extract was able to significantly reduce

blood sugar levels, whereas at the EKH 100 and 200 mg/ KgBW doses it was able to reduce blood sugar levels to reach the target below 200 mg / KgBW.

Table 2 shows the levels of GLUT4 in muscle tissue, where the GLUT4 protein is an important transporter in the regulation of glucose intake into cells. Increased expression of GLUT4 in a tissue indicates an increase in the ability of cells to intake glucose. In white rats induced with diabetes mellitus, there was a decrease in GLUT4 levels in muscle tissue. The administration of metformin drugs or sambiloto extracts has shown the ability to increase GLUT4 protein levels.

Table 3 shows the secondary metabolite content of the combined extract. The combined extract is rich in flavonoids. The dominant flavonoids in bay leaf extract are believed to be responsible for the effect of blood glucose regulation.

The content of flavonoids is the main secondary metabolite compound that is believed to play a role in blood glucose regulation.⁷⁻¹¹ Flavonoids increase the expression of GLUT 4 protein in muscle tissue. Increased expression of GLUT 4 causes an increase in glucose intake in cells. Increased glucose intake into cells, causes a decrease in the buildup of glucose outside the cells and interstitial tissue, which leads to a decrease in blood glucose levels.¹²⁻¹⁷

No.	Group	Blood Glucose $(mg/dL) \pm SD$	P-Value*	
1.	Control	28.26 ± 3.41	0.00	
2.	Diabetes	496.23 ± 15.43	-	
3.	Diabetes + Met	97.41 ± 7.21	0.00	
4.	Diabetes + EKH 50	379.12 ± 21.43	0.00	
5.	Diabetes + EKH 100	279.11 ± 18.65	0.00	
6.	Diabetes + EKH 200	163.83 ± 10.12	0.00	

Table 1. Level of blood glucose in serum

^{*} VS Diabetes + Met; ANOVA, post hoc Bonferroni; p<0.05

Table 2. Level of GLUT 4 in muscle

No.	Group	GLUT4 (pg/mL) ± SD	P-Value*
1.	Control	228.26 ± 3.41	0.00
2.	Diabetes	56.23 ± 15.43	-
3.	Diabetes + Met	147.41 ± 7.21	0.00
4.	Diabetes + EKH 50	86.12 ± 21.43	0.00
5.	Diabetes + EKH 100	108.11 ± 18.65	0.00
6.	Diabetes + EKH 200	145.83 ± 10.12	0.00

^{*} VS Diabetes + Met; ANOVA, pos hoc Bonferroni; p<0.05

Table 3. Phytochemical test

Material test	Saponins	Alkaloids	Triterpenoids	Steroids	Flavonoids
ЕКН	+	-	+	++	++++

4. Conclusion

The herbal combination extract has an effect on lowering blood sugar levels in Diabetes Mellitus White Rats by increasing glucose intake in cells and tissues.

5. References

- Asgary S, Rahimi P, Mahzouni P, Madani H. Antidiabetic effect of hydroalcoholic extract of Carthamus tinctorius L in alloxan-induced diabetic rats. J Res Med Sci. 2012; 17: 386
- Brownlee M. Biochemistry and molecular cell biology of diabetic complications. Nature. 2001; 414: 813–20.
- 3. Dey L, Attele AS, Yuan CS. Alternative therapies for type 2 diabetes. Altern Med Rev. 2002; 7: 45–58.
- Eddouks M, Lemhadri A, Michel JB. Hypolipidemic activity of aqueous extract of Capparisspinosa L in normal and diabetic rats. J Ethnopharmacol. 2005; 98: 345–50.
- 5. Eddouks M, Lemhardi A, Micel JB. Caraway and caper: potential antihyperglycaemic plants in diabetic rats. J Ethnopharmacol.

2004; 94: 143- 148.

- Fabiane K, Ricardo S, Oliveira T, Nagem TJ, Pinto AD, Oliveira MG, Soares JF. Effect of flavonoids morin; quercetin and nicotinic acid on lipid metabolism of rats experimentally fed with triton. Braz Arch Biol Techn. 2001; 44: 263–267.
- Grover JK, Yadav S, Vats V. Medicinal plants of India with anti-diabetic potential. J Ethnopharmacol. 2002; 8: 81–100.
- González-Villalpando C, López-Ridaura R, Campuzano JC, González-Villalpando ME. The status of diabetes care in Mexican population: Are we making a difference? Results of the National Health and Nutrition Survey 2006. Salud Publica Mex. 2010; 52: S36–46.
- Inzuchi SE, Maggs DG, Spollett GR, Page SL, Rite FS, Walton V. Efficacy and metabolic effect of Metformin and troglitazon in type II diabetes mellitus. N Engl J Med. 1998; 338: 867–872.

- Khan A, Anderson RA. Insulin potentiating factor (IPF) present in foods, species and natural products. Pak J Nutr. 2003; 2: 254– 257.
- 11. Khanfar MA, Sabri SS, Zarga MH, Zeller KP.
 The chemical constituents of Capparis spinosa of Jordanian origin. Nat Prod Res. 2003;17: 9–14.
- 12. Matsuyama T, Shoji K, Takase H, Kamimaki I, Tanaka Y, Otsuka A, et al. Effects of phytosterols in diacylglycerol as part of diet therapy on hyperlipidemia in children. Asia Pac J Clin Nutr. 2007; 16: 40–48.
- 13. Matthaus B, Ozcan M. Glucosinolates and fatty acid, sterol, and tocopherol composition of seed oils from Capparis spinosa Var spinosa and Capparis ovata Desf. Var canescens (Coss.) Heywood. J Agric Food Chem. 2005; 53: 7136–7141.
- 14. Moon J1, Lee SM, Do HJ, Cho Y, Chung JH, Shin MJ. Quercetin up-regulates LDL Receptor Expression in HepG2 Cells. Phytother Res. 2012; 26: 168–1694.
- 15. Nesto R. CHD: a major burden in type 2 diabetes. Acta Diabetol. 2001; 38: 3-8.
- 16. NRC (National Research Council) Institute of Laboratory Animal Resources, Commission on Life Sciences. Guide for the Care and Use of Laboratory Animals. Washington, D.C: National Academy Press; 1996.
- 17. Samarghandian S, Hadjzadeh M, Amin Nya F, Davoodi S. Antihyperglycemic and antihyperlipidemic effects of guar gum on streptozotocin-induced diabetes in male rats. Pharmacogn Mag. 2012; 8: 65–72.